

EXPLORING SR CALCIUM AND CYTOSOLIC CALCIUM WAVE DYNAMICS USING A 3D STOCHASTIC MYOCYTE MODEL

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INTRODUCTION

CALCIUM WAVES - EXPERIMENT

The local regulation of intracellular calcium has been widely known to play an important role in the normal excitation-contraction coupling in the cardiac myocytes. While Ca²⁺ sparks do not normally trigger regenerative SR Ca²⁺ release (i.e. Ca²⁺ waves), under calcium overload conditions calcium sparks can trigger spontaneous calcium waves. While experimental imaging using confocal microscopy with fluorescent dyes is the major technique used to study calcium dynamics inside a cardiac myocyte, the dynamics of SR calcium during calcium overload is poorly understood and controversial. In the study presented here, we present a new temporal-spatial model of calcium sparks to examine this issue. The resolution is sufficiently high (100nm) so that all critical local fluxes can be properly considered. The model also takes into account the different expression levels of the sodiumcalcium exchanger (NCX) and SERCA pumps near the SR Ca²⁺ release sites. From that, we want to demonstrate different factors contributing to the characteristics of Ca²⁻ sparks. This work includes an examination of the extent of SR calcium elevation, and how much calcium must be released to support the triggering of a sustained propagating Ca2+ wave within the cell. The findings presented here will be compared to published work in other studies.



Fig. 1: Calcium waves under $[Ca]_o = 5mM$ overload condition, the repetitive waves occur at a particular sites for each cell. This suggest there is a more density of release sites surrounding that region that allows mass calcium release is high enough to trigger the wave. Some waves can sustain to the next end, while some decay and stop in between, which suggests a stochastic nature of the waves.

CALCIUM SPARKS



Fig. 4: (A) A simulated calcium sparks, **(B)** Free calcium shows the underlying structure of release site (the delayed activation of the two satellite clusters are invisible under fluorescence profile, **(C)** The profile of a calcium spark giving FWHM=1.85um (each color represents the snapshot at different time point after the peak (e.g. bk=0 means black line at 0ms delayed); **(D)** The free calcium profile using back-calculation method agrees with experimental estimates, however, it underestimates the real free myoplasmic calcium amplitude.

DYNAMICS OF SR CALCIUM DEPLETION WAVES



CONCLUSIONS

- The spatio-temporal model with realistic CRU distribution, CRU geometry suggests the following:
- The spark model can reproduce realistic spark characteristics (FWHM=1.85µm, duration, rise time) by using realistic RyR dynamics, buffering, geometry, and diffusion.
- The back-calculation method underestimate s the true free calcium.
- The non-uniform placement of CRU is important for calcium wave initiation and propagation. Under calcium overload.

ACKNOWLEDGMENTS

This work is supported by the National Institutes of Health Grants 5R01HL105239 and 5R01AR057348, and a grant from the NVIDIA Corporation. The research leading to the results has received funding from the European Community's Seventh Framework Programme FP7/2007-2013 under grant agreement No HEALTH-F2-2009-241526, EUTrigTreat.

3D WHOLE-CELL RAT MODEL

3D Temporal-Spatial Cell Model Rat ventricular myocyte: Cell dimension: 100x20x18 (µm³) (V_{cell}=36 pL) • grid size: $0.2 \,\mu\text{m} \rightarrow N_{\text{mesh}} = 4,500,000$ \$ L_X mesh points V_{nsr}^T = 3.5% V_{cell}, V_{mvo}^T = 50% V_{cell} 20,000 CRUs/cell (B) Inter-CRU distances: non-uniform distribution of CRUs CRU MODEL Each CRU: A dyad subspace: 300x300x12 (nm³) A junctional SR: 300x300x50 (nm³) A network SR: · Cytoplasm Spark simulation: 36 RyR2 + 2 satellite clusters of RyR, each with 15RyR at 100nm distance ある • Ca²⁺ wave simulation: 49 RyR2 Ryanodine receptor-2 (RyR2) [2] 2-state minimal model · Stochastic gating, with allosteric coupling to each other 1 Both cytosolic Ca²⁺ sensitivity and luminal Ca²⁺ regulation.

Fluxes

- J_{efflux} : from subspace
- J_{refill} : from local NSR
- J_{ryr}: from JSR to subspace via opening RyR2s





Fig. 2: The placement of calcium release sites **(A)** at one z-depth, **(B)** at one Z-disc. The distribution of inter-CRU distance is derived based on the experimental



80	(1114.65ms)	20	40	60. Distance (μm)	
80		20	40	60 Distance (µm)	
80	0 (1424.21ms)	20	40	60 Distance (µm)	
60	(1692.51ms)	20	40	60 Distance (µm)	80
80	(2068.40ms)	20	40	60 Distorce (µm)	

Fig. 3 Calcium overload ([Ca]_{nsr}=1.7mM, [Ca]_i=0.15 μ M), this computational model of the rat ventricular myocyte can reproduce a repetitive sustained calcium wave which typically initiates at one end of the cell. The initiation site typically occurs where release sites are closer together or at a boundary.

(2382.58ms)