### **IMAG Theme 3**

Top Down Mesoscale Simulations and Free Energy Calculations: Experimentally Validated (Minimal) Models for Receptor Trafficking and Nanocarrier Adhesion

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# **Physically Based Multiscale Models**



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# **Minimal Models for Intracellular Trafficking**



### Predictive Biophysical Models for Protein-Mediated Membrane Processes at the Mesoscale



## **Mesoscale Elastic Model for Membranes**

### Helfrich Free Energy Nelson, Piran, Weinberg, 1987

$$f_{\rm c} = \frac{1}{2}k\left(H - H_0\right)^2 + \bar{k}K.$$

H<sub>0</sub>:Intrinsic curvature k: Bending Modulus k: Gaussian Curvature Modulus



$$E = \int_{A} \frac{\kappa}{2} (H - H_0)^2 dA + \sigma (A - A_{\text{flat}})^2$$

Elastic free energy including frame tension on a membrane patch

 $\begin{array}{l} H \Rightarrow 1/2[1/R_1 + 1/R_2] \\ K \Rightarrow 1/R_1 \times 1/R_2 \end{array}$ 

Plane: H=0, K=0



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# Coarse-Grained Representation of Protein-Membrane Interaction



Integral Membrane Proteins

Membrane is attached to the protein at a fixed contact angle Goulian M, Bruinsma R, Pincus P (1993) Lubensky T (1997) Kim KS, Neu J, Oster G (1998)

#### LOCAL CURVATURE INDUCER MODEL

#### McMahon, 2003, 2005

Tubule diameter=20 nm;  $\Delta E_{\text{binding}}$ = -14 k<sub>B</sub>T





Seifert, et. al. 2006; Weinstein, Radhakrishnan, 2006; Agrawal, Weinstein, Radhakrishnan, 2008  $C_0=0.1 \text{ nm}^{-1}$ ; b=8 nm;  $\kappa=20 \text{ k}_B\text{T}$ b nm



$$H_0 = C_0 e^{-s^2/b^2}$$



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## **Generalized Langevin Dynamics**

Helfrich Hamiltonian for fluid lipid membrane [Helfrich (1973) *Z. Naturforsch* **28c** 693]

$$E = \int_{S} \left[ \frac{\kappa}{2} (H - H_0)^2 + \overline{\kappa} K \right] dA \qquad \begin{array}{c} H = c_1 + c_2 & \text{Mean curvature} \\ K = c_1 c_2 & \text{Gaussian curvature} \end{array}$$

*k*: bending rigidity *k*: Gauss curvature modulus  $H_0$ : instantaneous mean curvature Monge (linear) model [Agrawal *et al.* (2008) *Mol. Phys.*]

$$z = z(x, y) \quad A_{ij} = A_{flat,ij} \left[ 1 + (\nabla z)^2 \right]^{\frac{1}{2}}$$

$$H \approx z_{xx} + z_{yy} = \nabla^2 z$$

$$E = \iint_{R} \left[ \frac{\kappa}{2} (\nabla^2 z - H_0)^2 + \frac{\kappa}{4} H_0^2 (\nabla z)^2 \right] dx dy$$

$$\frac{\partial z(r,t)}{\partial t} = -M \frac{\delta E}{\delta z} + \xi(r,t)$$

$$\stackrel{M: \text{ mobility term}}{\xi: \text{ thermal noise term}}$$

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Curvatures are calculated through coordinate transformation between global and local Darboux frame. One of the two moves is randomly selected and the energy change is calculated.





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### **Protein-Mediated Membrane Fluctuations**

Weinstein, Radhakrishnan, Mol Phys, 2006; Agrawal, Radhakrishnan, Mol Phys, 2008



Proteins perform a random walk on membrane surface with a membrane mediated force field



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## Membrane-Mediated Potential of Mean Force (PMF) between Proteins

- PMF is dictated by both energetic and entropic components
- Energy: Epsin experience repulsion due to energetic component when brought close.
- Entropy:

$$\delta^{2} E(\phi) = \iint_{A} \kappa \left( \nabla^{2} \phi \right)^{2} + \left( \frac{\kappa}{2} H_{0}^{2} + \sigma \right) \left( \nabla \phi \right)^{2} dx dy > 0$$

 $\delta^2 E$ ~ spring constant;  $\phi$ =test function

- Regions of non-zero H<sub>0</sub> assume increased stiffness and hence reduced membrane fluctuations
- The system can lower its free energy by localizing epsins on the membrane



### **Need for Free Energy Calculations**



**Department of Bioengineering** 

#### Agrawal, Radhakrishnan, Mol Phys, 2008

## Free Energy Calculations via Thermodynamic Integration



Free energy change with spatial extent of the curvature function





Entropy change is small (<5% of the Energy change) but is of order  $k_BT$ 

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Agrawal, Radhakrishnan, PRE, 2009

$$\left(\frac{\partial F}{\partial \lambda}\right)_{N,V,T} = -\frac{1}{\beta} \frac{\partial}{\partial \lambda} \ln Q = \left\langle \frac{\partial E}{\partial \lambda} \right\rangle_{\lambda}$$

By choosing  $\lambda = C_0$ ,

$$\frac{\partial F}{\partial C_0} = \left\langle \Gamma(r_0) \kappa \sum_{N} \left[ -\left( \nabla^2 z_i - \lambda \Gamma(r_0) \right) + \frac{\lambda}{2} \left( \nabla z_i \right)^2 \right] \left( \Delta r_i \right)^2 \right\rangle$$

$$\Delta F = F(C_0) - F(0) = \int_0^{C_0} \frac{\partial F}{\partial C_0} dC_0$$

# Vesicle Nucleation in Clathrin-Mediated Endocytosis





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## Bond-Orientational Patterning of Epsin on Clathrin Lattice Leads to a Mature Vesicle Formation



[Agrawal, Radhakrishnan, Plos Comput Biol, 2010]



### Receptor Trafficking: Bioenergetics of Clathrin Induced Membrane Vesiculation

Agrawal, Radhakrishnan, Plos Comput Biol, 2010 Ramanan et al, Integrative Biology, 2011



Weak hydrophobic interactions lead to self assembly of clathrin coat Epsin-Induced Curvature Stabilizes Mature Vesicular Intermediates



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### Free Energy Considerations in Clathrin Induced Membrane Vesiculation Calculated Experimental



30 20

area of the coat [nm<sup>2</sup>]

Agrawal, Radhakrishnan, Plos Comp Biol, 2010

Control

10000



Jakobsson, J.; PNAS 2008, 6445.

Our results highlight the unique and central role played by epsin in the process of vesicle nucleation during endocytosis

15000



3

2

1

-2

-3Ľ 0

E/k<sub>B</sub>T

control

-CLAP laG

5000

Ensemble of Epsin-Membrane CGMD Simulations; each replica with a unique lateral pressure





**CG/CM** Interface for

**CGMD** gives curvature field



Mesoscale MC model gives positions and frame tension (lateral pressure)



Two-way multiscale coupling; epsinmembrane simulations in will quantify: 1.Curvature field 2.Curvature induction by multiple epsins 3.Local stress fields Model for predicting protein partitioning and vesicle budding events under various conditions: different proteins, cytoskeletal tension, ECM stiffness



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