## **IMAG Theme 3**

*Top Down Mesoscale Simulations and Free Energy Calculations: Experimentally Validated (Minimal) Models for Receptor Trafficking and Nanocarrier Adhesion*

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# **Physically Based Multiscale Models**





# **Minimal Models for Intracellular Trafficking**



## Predictive Biophysical Models for Protein-Mediated Membrane Processes at the Mesoscale



# **Mesoscale Elastic Model for Membranes**

## **Helfrich Free Energy Nelson, Piran, Weinberg, 1987**

$$
f_{\rm c} = \frac{1}{2}k\left(H - H_0\right)^2 + \overline{k}K.
$$

 $H_0$ :Intrinsic curvature k: Bending Modulus **k: Gaussian Curvature Modulus** 



$$
E = \int_{A}^{K} \frac{(H - H_0)^2 dA + \sigma (A - A_{\text{flat}})}{2}
$$

Elastic free energy including frame tension on a membrane patch

 $H \implies 1/2[1/R_1 + 1/R_2]$  $K\rightarrow 1/R_1 \times 1/R_2$ 

Hane:

2



### **Coarse-Grained Representation of Protein-Membrane Interaction** *H*<sub>0</sub> =∑ $Ce^{e^{s(fbsa)}}$



**Integral Membrane Proteins** Membrane is attached to the protein at a fixed contact angle Goulian M, Bruinsma R, Pincus P (1993) Lubensky T (1997) Kim KS, Neu J, Oster G (1998)

### LOCAL CURVATURE INDUCER MODEL

### McMahon, 2003, 2005

Tubule diameter=20 nm;  $\Delta E_{binding}$ = -14  $k_BT$ 





b nm  $C_0$ =0.1 nm<sup>-1</sup>; b=8 nm;  $\kappa$ =20 k<sub>B</sub>T Seifert, et. al. 2006; Weinstein, Radhakrishnan, 2006; Agrawal, Weinstein, Radhakrishnan, 2008

$$
\frac{1}{\mathcal{S}} = \frac{1}{\sqrt{\frac{1}{\mathcal{S}}\mathcal{S}}}
$$

$$
H_0 = C_0 e^{-s^2/b^2}
$$



# **Generalized Langevin Dynamics**

Helfrich Hamiltonian for fluid lipid membrane [Helfrich (1973) *Z. Naturforsch* **28c** 693]

$$
E = \int_{s}^{s} \left[ \frac{\kappa}{2} (H - H_0)^2 + \overline{\kappa} K \right] dA
$$
  
\n $H = c_1 + c_2$   
\n $H = c_1 c_2$   
\n**Mean curvature**  
\n**Gaussian curvature**

 $\kappa$ : bending rigidity  $\bar{\kappa}$ : Gauss curvature modulus  $H_0$ : instantaneous mean curvature

Monge (linear) model [Agrawal *et al.* (2008) *Mol. Phys.*]

$$
z = z(x, y) A_{ij} = A_{flat,ij} \left[ 1 + (\nabla z)^2 \right]^{\frac{1}{2}}
$$
  
\n
$$
H \approx z_{xx} + z_{yy} = \nabla^2 z
$$
  
\n
$$
E = \iint_R \left[ \frac{\kappa}{2} (\nabla^2 z - H_0)^2 + \frac{\kappa}{4} H_0^2 (\nabla z)^2 \right] dxdy
$$
  
\n
$$
\frac{\partial z(r,t)}{\partial t} = -M \frac{\delta E}{\delta z} + \xi(r,t)
$$
  
\n<sup>M</sup>: mobility term  
\n<sup>M</sup>: the final noise term  
\n<sup>M</sup>: the final noise term





Curvatures are calculated through coordinate transformation between global and local Darboux frame. One of the two moves is randomly selected and the energy change is calculated.





### Protein-Mediated Membrane Fluctuations

Weinstein, Radhakrishnan, Mol Phys, 2006; Agrawal, Radhakrishnan, Mol Phys, 2008



 Proteins perform a random walk on membrane surface with a membrane mediated force field



## **Membrane-Mediated Potential of Mean Force (PMF) between Proteins**

- **PMF** is dictated by both energetic and entropic components
- **Energy:** Epsin experience repulsion due to energetic component when brought close.

### **Entropy:**

$$
\delta^2 E(\phi) = \iint_A \kappa (\nabla^2 \phi)^2 + \left(\frac{\kappa}{2} H_0^2 + \sigma\right) (\nabla \phi)^2 dxdy > 0
$$

 $\delta^2$ E ~ spring constant;  $\phi$  = test function

- Regions of non-zero  $H_0$  assume increased stiffness and hence reduced membrane fluctuations
- **The system can lower its free energy** by localizing epsins on the membrane





**University of Pennsylvania Department of Bioengineering**

### x 10-15 Agrawal, Radhakrishnan, Mol Phys, 2008

# **Free Energy Calculations via Thermodynamic Integration**



Free energy change with spatial extent of the curvature





Entropy change is small (<5% of the Energy change) but is of order  $k_BT$ 

Agrawal, Radhakrishnan, PRE, 2009

$$
\left(\frac{\partial F}{\partial \lambda}\right)_{N,V,T} = -\frac{1}{\beta} \frac{\partial}{\partial \lambda} \ln Q = \left\langle \frac{\partial E}{\partial \lambda} \right\rangle_{\lambda}
$$

By choosing  $\lambda = C_0$ ,

$$
\frac{\partial F}{\partial C_0} = \left\langle \Gamma(r_0) \kappa \sum_N \left[ -\left(\nabla^2 z_i - \lambda \Gamma(r_0)\right) + \frac{\lambda}{2} \left(\nabla z_i\right)^2 \right] \left(\Delta r_i\right)^2 \right\rangle
$$

$$
\Delta F = F(C_0) - F(0) = \int_0^{C_0} \frac{\partial F}{\partial C_0} dC_0
$$





# **Vesicle Nucleation in Clathrin-Mediated Endocytosis**





# **Bond-Orientational Patterning of Epsin on Clathrin Lattice Leads to a Mature Vesicle Formation**



[Agrawal, Radhakrishnan, Plos Comput Biol, 2010]



### **Receptor Trafficking: Bioenergetics of Clathrin Induced Membrane Vesiculation**

Agrawal, Radhakrishnan, Plos Comput Biol, 2010 Ramanan et al, Integrative Biology, 2011



Weak hydrophobic interactions lead to self assembly of clathrin coat Epsin-Induced Curvature Stabilizes Mature Vesicular Intermediates



### **Calculated Experimental Free Energy Considerations in Clathrin Induced Membrane Vesiculation**



**Our results highlight the unique and central role played by epsin in the process of vesicle nucleation during endocytosis**



*Ensemble of Epsin-Membrane CGMD Simulations; each replica with a unique lateral pressure* 



**CG/CM Interface for Coupling CGMD: coarse- grained molecular dynamics with CM: continuum Helfrich MC**

*CGMD gives curvature field*



*Mesoscale MC model gives positions and frame tension (lateral pressure)*



*Two-way multiscale coupling; epsinmembrane simulations in will quantify: 1.Curvature field 2.Curvature induction by multiple epsins 3.Local stress fields*

*Model for predicting protein partitioning and vesicle budding events under various conditions: different proteins, cytoskeletal tension, ECM stiffness*

