A multi-scale modeling of the innate immune response to fungal respiratory pathogens

BACKGROUND

Invasive pulmonary aspergillosis is a severe pneumonia in immunocompromised hosts. It is caused by the inhalation of the spores of the saprophytic fungus Aspergillus, characterized by the growth of hyphae within the tissues of the lung. Iron acquisition is a critical component of microbial pathogenesis that is essential to microbial virulence. An important component of the innate immune response is to sequester iron from the pathogen, an instance of nutritional immunity, which involves several processes at different spatial and temporal scales, ranging from molecular events inside several types of immune cells, tissue-level events, and involvement of the liver. As fungal pathogens have evolved mechanisms to actively acquire iron from the host, a "battle for iron" plays out in the very early stages of infection. Our hypothesis is that host-centric interventions can result in a favorable outcome of this battle.



Figure 1: A Simplified description of the model. *Aspergillus* induces macrophages (MØ) and epithelial cells to produce chemokines and cytokines that attract neutrophils (N); neutrophils and macrophages kill Aspergillus; Aspergillus secretes the siderophore TAFC that chelates iron from transferrin bound to iron (TfBI); macrophages secrete IL-6 which instructs the liver to secrete hepcidin; hepcidin reduces the level of serum transferrin (Tf) and inhibits macrophage iron export. Neutrophil recruitment and killing of *Aspergillus* is more pronounced than macrophage (thick arrows). Red arrows represent iron-related actions.

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Figure 2: Model validation with data from the literature [1,2]. Simulated data were obtained from model parameters in the literature not including these two papers. Predicted quantity of leukocytes (macrophages and neutrophils) 24 hours post-infection was compared with quantities reported in the literature (left). Predicted lung concentration of TNF-a and MIP-1b 24 hours postinfection were compared with data from the literature (right). Dark bar predicted, light bar reported in the literature. Error bars represent standard deviation.

Table 1: Sensitivity analysis performed with partial ranking correlation coefficient and Latin-hypercube-sampling. The table shows the mean plus or minus standard deviation. The table presents the parameters that were more than 1.96 standard deviations away from zero. In bold are the three top parameters by absolute correlation from each model. Parameters were subdivided into four groups: recruitment and chemotaxis (orange), iron-related (red), strength of the immune response (blue), and fungal biology (green). Notice that, in general, the highest partial correlations were related to recruitment and chemotaxis. But there were also many parameters related to iron immune response.



Mathematical modeling can be an effective approach to the exploration of mechanisms of innate immunity in the lung in response to fungal pathogens. It allows the integration of known biophysics, biology, and immunology, as well as experimental data into a multi-scale computational model that can serve as an investigative tool to guide experiments.

RESULTS

Figure 3: Sample simulation run. WT and neutrophil KO. WT evolves to clearance while KO shows unrestrained growth. The scale is in log base 10 for better visualization. Unbounded growth is to be interpreted as a failure of the innate immune response to control the infection.

Im	Neutropenic	Description	
-0	-0.5153 ± 0.0376	global recruitment rate	
-	$\textbf{-0.4404} \pm \textbf{0.0409}$	MIP-1b secretion rate	
-0	-0.0006 ± 0.0503	MIP-2 secretion rate	
0	0.1474 ± 0.0459	global diffusion rate	
_	0.1654 ± 0.0469	TAFC uptake rate	
	0.1793 ± 0.0457	TAFC secretion rate	
-	-0.1253 ± 0.0511	Michaelis constant of transferrin-TAFC reaction	
	-0.2096 ± 0.0465	A. fumigatus metabolic sensibility to iron	
_	-0.1765 ± 0.0515	slope of the function hepcidin-transferrin	
_	-0.0720 ± 0.0468	macrophage phagocytosis probability	
_	0.0045 ± 0.0512	bility to interact with hyphae (macrophage and neutrophil)	
_	-0.2661 ± 0.0488	probability to kill internalized conidia	
	0.4345 ± 0.0402	time cells remain activate without stimulus	
	-0.1646 ± 0.0502	inverse of growth rate	
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CONCLUDING REMARKS

